

Fall 9-9-2014

HO-1 Upregulation Attenuates Adipocyte Dysfunction, Obesity, and Isoprostane Levels in Mice Fed High Fructose Diets

Zeid Khitan

Marshall University, zkhitan@marshall.edu

Mohit Harsh

Komal Sodhi

Joseph I. Shapiro MD

Marshall University, shapiroj@marshall.edu

Nader G. Abraham

Marshall University, abrahamn@marshall.edu

Follow this and additional works at: http://mds.marshall.edu/sp_psr

 Part of the [Medicinal and Pharmaceutical Chemistry Commons](#), and the [Other Pharmacy and Pharmaceutical Sciences Commons](#)

Recommended Citation

Khitan Z, M Harsh, K Sodhi, JI Shapiro and NG Abraham. 2014. HO-1 upregulation attenuates adipocyte dysfunction, obesity, and isoprostane levels in mice fed high fructose diets. *J. Nutr. Metab.* Vol. 2014:1-13. PMID:25295182.

This Article is brought to you for free and open access by the Faculty Research at Marshall Digital Scholar. It has been accepted for inclusion in Pharmaceutical Science and Research by an authorized administrator of Marshall Digital Scholar. For more information, please contact zhangj@marshall.edu.

Research Article

HO-1 Upregulation Attenuates Adipocyte Dysfunction, Obesity, and Isoprostane Levels in Mice Fed High Fructose Diets

Zeid Khitan,¹ Mohit Harsh,¹ Komal Sodhi,¹ Joseph I. Shapiro,¹ and Nader G. Abraham^{1,2}

¹ Department of Internal Medicine, Marshall University Joan C. Edwards School of Medicine, Huntington, WV 25701, USA

² Departments of Medicine and Pharmacology, New York Medical College, Valhalla, NY 10595, USA

Correspondence should be addressed to Nader G. Abraham; nader_abraham@nymc.edu

Received 19 June 2014; Accepted 14 August 2014; Published 9 September 2014

Academic Editor: Stan Kubow

Copyright © 2014 Zeid Khitan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Fructose metabolism is an unregulated metabolic pathway and excessive fructose consumption is known to activate ROS. HO-1 is a potent antioxidant gene that plays a key role in decreasing ROS and isoprostanes. We examined whether the fructose-mediated increase in adipocyte dysfunction involves an increase in isoprostanes and that pharmacological induction of HO-1 would decrease both isoprostane levels and adipogenesis. **Methods and Results.** We examined the effect of fructose, on adipogenesis in human MSCs in the presence and absence of CoPP, an inducer of HO-1. Fructose increased adipogenesis and the number of large lipid droplets while decreasing the number of small lipid droplets ($P < 0.05$). Levels of heme and isoprostane in fructose treated MSC-derived adipocytes were increased. CoPP reversed these effects and markedly increased HO-1 and the Wnt signaling pathway. The high fructose diet increased heme levels in adipose tissue and increased circulating isoprostane levels ($P < 0.05$ versus control). Fructose diets decreased HO-1 and adiponectin levels in adipose tissue. Induction of HO-1 by CoPP decreased isoprostane synthesis ($P < 0.05$ versus fructose). **Conclusion.** Fructose treatment resulted in increased isoprostane production and adipocyte dysfunction, which was reversed by the increased expression of HO-1.

1. Introduction

Over the past several decades the use of fructose, as a sweetener, has emerged in processed foods, soft drinks, and an assortment of desserts [1–3]. Fructose has been linked to the development and exacerbation of centripetal obesity and metabolic syndrome, in both animals and humans [3]. In addition, excessive fructose consumption (>50 g/d) is involved in the development of metabolic imbalance and diabetes, independent of caloric restriction and weight gain [2, 4]. Sucrose, commonly found in table sugar, is a disaccharide comprising equimolar amounts of fructose and glucose and is the worldwide primary source for dietary fructose [1]. In light of the correlation between fructose and obesity, it is imperative to examine the pathogenesis by which fructose may lead to metabolic syndrome and adiposity.

Fructose is absorbed by the small intestine via passage through GLUT 5 transporters and is metabolized through the glycolytic pathway [1, 5]. Excessive fructose intake is

associated with elevated levels of inflammatory mediators and reactive oxygen species (ROS), thereby contributing to the onset of metabolic syndrome and obesity [6]. Obesity is associated with oxidative stress, increased levels of ROS, and an increase in free heme, a known prooxidant [7, 8].

During oxidative stress, heme is released from cells and tissue and contributes to the growing “free heme pool.” Free heme is a potent prooxidant and increases ROS levels [8–11]. Elevated levels of heme are toxic to the body and catalyze oxidative damage to cells, tissues, and organs. Obesity and oxidative stress increase cellular heme and the heme-mediated increased generation of isoprostanes [8, 10, 11]. Isoprostanes are prostaglandin-like compounds formed *in vivo* from the free radical-catalyzed peroxidation of the essential fatty acid, arachidonic acid [12]. These nonclassical eicosanoids have strong biological activity as inflammatory mediators [13] and are recognized as markers of lipid peroxidation in both animal and human models of oxidative stress [11, 14, 15]. In obese patients, urine isoprostane levels increase

with increasing BMI in both male and female subjects [16]. Plasma concentration of isoprostane increases during acute fructose loading [17]. Fructose-induced oxidative stress and insulin resistance in humans are prevented by grape polyphenols [18].

Excessive fructose consumption leads to the elevation of ROS [19] and may lead to increased lipid peroxidation, altered metabolic homeostasis, and hypertrophic adipogenesis. Adipogenesis begins with the commitment of MSCs to the adipocyte lineage, followed by terminal differentiation of preadipocytes to mature adipocytes [20, 21]. Fat tissue derived adipocytes express several regulatory proteins such as Wnts and β -catenin, which repress adipogenic differentiation in preadipocytes [22, 23].

Obesity increases oxidative stress and concurrently decreases expression and activity of key cytoprotective enzyme systems, including heme oxygenase-1 (HO-1) [24, 25]. Heme oxygenase-1 is a stress-response protein and decreased expression results in an increase in cellular heme content, resulting in oxidative stress and adipocyte dysfunction [7, 26]. Induction of HO-1 decreases endogenous ROS and the protective effect of increased HO-1 expression is ascribed to several factors that include a reduction in cellular heme, the induction of the iron binding protein, ferritin, and the increased formation of bilirubin, which efficiently scavenges toxic oxygen species [7]. Induction of HO-1 expression *in vivo* and *in vitro* is associated with an increase in preadipocytes, a reduction in the number of enlarged adipocytes, elevated adiponectin secretion, and an increase in the number of small adipocytes, which are regarded as "healthy" adipocytes [27–31]. More importantly, heterozygous HO-1 knockout mice treated with an inducer of HO-1 failed to increase the production of nitric oxide [32]. Improvement in nitric oxide bioavailability and vascular function are critical to arterial induction of HO-1 [32].

The goal of this study was to determine the role of HO-1 gene expression in the fructose-mediated increase in adiposity, inflammation, and metabolic imbalance, in *in vitro* and *in vivo* models of excessive fructose intake. This is the first study to demonstrate that increased expression of HO-1 plays a significant role in modulating the phenotype of adipocytes via a decrease in heme and isoprostane levels in MSC-derived adipocytes treated with fructose and mice fed a high fructose diet.

2. Materials and Methods

2.1. Experimental Design for *In Vitro* Experiment

2.1.1. Human Bone Marrow-Derived MSC Culture. Frozen bone marrow mononuclear cells were purchased from All-cells (Emeryville, CA). After thawing, mononuclear cells (MSCs) were resuspended in α -minimal essential medium (α -MEM, Invitrogen, Carlsbad, CA) that was supplemented with 20% FBS and 1% A/A. The cells were transferred into a 75 cm² flask, cultured in α -MEM with supplements, and maintained at 37°C in a 5% CO₂ incubator. Medium was changed after 24 hours and every 2 days thereafter. Once 80%

confluent, the MSCs were recovered by the addition of 0.25% trypsin/EDTA, split, and partitioned into additional 75 cm² flasks.

2.1.2. Human Bone Marrow-Derived MSC Adipocyte Differentiation. MSCs (passage 2-3) were plated in 75 cm² flasks at a density of 1-2 \times 10⁴ cells per cm² and were again cultured in their respective maintenance media until cells were 80% confluent. The maintenance medium was then replaced with adipogenic medium and the cells were cultured for an additional 14 days. The adipogenic medium consisted of complete culture medium supplemented with DMEM (Invitrogen, Carlsbad, CA), 10% FBS, 1% A/A, 10 μ g/mL insulin, 1 μ M dexamethasone (Sigma-Aldrich, St. Louis, MO), and 0.1 mM indomethacin (Sigma-Aldrich, St. Louis, MO). Human MSCs were cultured in this adipogenic differentiation medium along with fructose in the presence and absence of the HO-1 inducer, cobalt protoporphyrin (CoPP) (5 μ M), and the HO activity inhibitor, tin (Sn⁴⁺)-mesoporphyrin (SnMP) (5 μ M). Medium and treatments were renewed daily. Concurrently, human bone marrow-derived MSCs were plated in 24-well plates at a density of 1 \times 10⁴ cells per cm² and maintained for 24 hours in their respective maintenance media. Following this initial 24 hours, cells were treated with the aforementioned adipogenic differentiation medium in conjunction with fructose in the presence and absence of CoPP (5 μ M) and SnMP (5 μ M) for 14 days. MTT assay was done to determine the influence of all treatments on cell viability and no significant change was seen between the groups (data not shown).

2.2. Experimental Design for *In Vivo* Experiment. All animal studies were approved by the Marshall University Animal Care and Use Committee in accordance with the National Institutes of Health Guidelines for Care and Use of Laboratory Animals. Forty eight-week-old C57Bl6 male mice were used in the studies. Mice were divided into 4 groups: 6 mice/group, control animals, mice fed a high fructose diet, and mice fed a high fructose diet and treated with CoPP, in the absence or presence of SnMP. Mice were fed either a normal chow diet containing 11% fat, 62% carbohydrate, and 27.0% protein with total calories of 12.6 KJ/g or a high fructose diet containing 60% fructose, 13.6% starch, 7% total fat, and 19.4% protein. The mice had free access to water. After four months of high fructose diet, the average weight of the fructose group showed a 1.4-fold increase in body weight compared to control animals. Administration of CoPP reversed the effect of fructose to the levels of animals fed regular chow (2 mg/100 g body weight) (4 mg/100 g bodyweight for the last month). Concurrent administration of SnMP resulted in an increase in body weight compared to mice fed fructose diets alone. We monitored food intake in mice receiving a normal chow diet or a high fructose diet and food intake did not change in any treatment group including those administered CoPP-SnMP. Blood samples were collected in K₃EDTA tubes at sacrifice and the plasma was separated. Samples were flash frozen in liquid nitrogen and maintained at -80°C until assayed.

2.3. Effect of Varying Fructose Concentrations on Adipogenesis. To determine the optimal effect of fructose on MSC-derived adipocyte differentiation, cells were treated with fructose at concentrations of 5, 50, and 500 μM . After 14 days, cells were stained with Oil Red O solution. Optimum concentration for fructose was found to be 500 μM and this concentration was utilized during treatment of cells throughout the experimentation.

2.4. Oil Red O Staining. 0.21% Oil Red O in 100% isopropanol (Sigma Aldrich, St. Louis, MO) was used for staining. Briefly, adipocytes were fixed in 4% formaldehyde, stained with Oil Red O for 10 min, and rinsed with pH 7.2 PBS (1x) (Invitrogen, Carlsbad, CA). Pictures were captured using an Olympus IX81 motorized inverted microscope. Oil Red O was eluted by adding 100% isopropanol for 10 min and OD was measured at 490 nm 24 hours later.

2.5. Measurement of Lipid Droplet Size. Cell size was measured using *ImagePro Analyzer* (Media Cybernetics, Inc., MD). The classification of the size of lipid droplets was based on size by area (pixels).

2.6. Detection of Isoprostane, Heme, and Cytokine Levels for In Vitro and In Vivo Experiments. Isoprostane levels were determined in conditioned media and in mouse plasma using an ELISA assay (Cayman Chemical, Ann Arbor, MI). Heme content in MSCs and mouse adipose tissue was determined by the pyridine hemochromogen method as described previously [33, 34]. The absorbance difference between λ 557 and λ 530 nm was used to calculate hepatic heme using an extinction coefficient of 20.7 $\text{mM}^{-1} \text{cm}^{-2}$. TNF α , IL-1 β , and IL-6 levels were determined in conditioned media and mouse serum using an ELISA assay (Pierce Biotechnology, Woburn, MA).

2.7. Preparation of Adipocyte Matrix from Adipose Tissues Using Collagenase. Subcutaneous and visceral fat visible in the abdomen mesenteric fat, fat around the liver, kidney, and spleen was dissected free at the time of sacrifice and used to extract fat cells for RNA. Fat tissue was minced and incubated in 20 mL of Hank's balanced salt solution (HBSS) 1x (without calcium or magnesium), buffer containing 1% BSA and 0.1% collagenase at 1-2 gm of fat/10 mL. The fat tissues were placed in 15 mL conical tubes containing RNA inhibitors for RNA extraction and incubated in a warm water bath for 2 minutes. Wash fat in 3 serial dilutions of Betadine solution in PBS and then mince fat with sterile scissors for 2-3 minutes followed by RNA.

2.8. RNA Extraction and Real-Time PCR for In Vitro and In Vivo Experiments. Total RNA was extracted from mice adipose tissue using RNeasy Protect Mini kit (QIAGEN, Maryland, USA) according to manufacturer's instructions. Total RNA (1 μg) was transcribed into cDNA using GeneAmp kit (Applied Biosystems, Branchburg, NJ, USA) reverse transcription reagents. Total RNA was analyzed by a quantitative real-time polymerase chain reaction (qRT-PCR). Real-time

PCR was performed using SYBR Green PCR Master Mix (Applied Biosystems) on a 7500 HT Fast Real-Time PCR System (Applied Biosystems). Specific primers for human MSCs were the following, with actin being used as an endogenous control described in Table 1. Specific primers for mouse tissue were as in Table 1, with actin being used as an endogenous control.

2.9. Statistical Analyses. Statistical significance was determined using one-way analysis of variance followed by Tukey-Kramer post hoc test. $P < 0.05$ was considered to be significant. Data are expressed as means \pm SEM.

3. Results

3.1. Fructose Increases Adipogenesis and Isoprostane Release in a Dose-Dependent Manner in MSCs. In order to determine the optimal concentration for adipocyte differentiation in cultured human MSCs, cells were treated with fructose, 5, 50, and 500 μM . This resulted in an increase in large lipid droplets in MSCs and a decrease in small lipid droplets in a dose-dependent manner (Figures 1(a) and 1(b)). 500 μM of fructose was determined to be the optimal concentration for stimulating adipogenesis ($P < 0.05$ versus control) (Figures 1(a) and 1(b)). Isoprostane levels increased in a dose-dependent manner in conditional media of cultured MSCs, with 50 and 500 μM , the concentration at which there was the significant increase in isoprostane levels compared to control as determined by ELISA (Figure 1(c), $P < 0.05$ versus control).

3.2. Isoprostane Release and Heme Content Increase during Adipogenesis. We examined if a relationship with adipogenesis and isoprostane and cellular heme levels existed in MSCs. We observed a significant increase in isoprostane levels from day 0 to day 14 (Figure 2(a)) along with an increase in cellular heme content (Figure 2(b)). These results show, for the first time, an increase in isoprostane production and heme content with fructose-mediated adipogenesis.

3.3. HO-1 Overexpression Attenuates Fructose-Mediated Adipogenesis and Lipid Accumulation. In order to examine the effect of overexpression of HO-1 on adipogenesis, we treated cells with CoPP with or without SnMP in the presence of 500 μM fructose (Figure 3). After 14 days, the number of lipid droplets stained with Oil Red O was lower after CoPP when compared to control (Figure 3(a), $P < 0.05$). In contrast, SnMP resulted in higher levels of Oil Red O staining when compared to control ($P < 0.05$). CoPP decreased the number of large lipid droplets and increased the number of small lipid droplets when compared to control (Figures 3(b) and 3(c), $P < 0.05$). SnMP reversed these effects when compared to both CoPP and control ($P < 0.05$).

3.4. HO-1 Overexpression Reduces Isoprostane Production and Adipogenic Marker Expression and Concurrently Increases Canonical Wnt Signaling in Human MSCs. To verify that fructose-mediated adipocyte dysfunction can be reversed via

TABLE 1: The sequence of primers used for real-time PCR.

Primer	Forward	Reverse
Actin	5'-AGGCACCAGGGCGTGAT-3'	5'-GCCCACATAGGAATCCTTCTGAC-3'
HO-1	5'-TGAAGGAGGCCACCAAGGAG-3'	5'-CCCCTGAGAGGTCACCCAGG-3'
Mest	5'-CCAGCACATCCCGGTGCTT-3'	5'-TTCCATGAGTGCAGAGCAGG-3'
β -Catenin	5'-GGCAGGATACACGGTGCCGC-3'	5'-CTTCAGGTACCCCTCAGGCCCGC-3'
Wnt10b	5'-GGTCTCGGCCTCCGCCCTTA-3'	5'-ACGAAACCGCGCTTGAGGA-3'
Adiponectin	5'-GACGACACCAAAAGGGCTCAGG-3'	5'-CATCACGGCCTGGTGTGCCA-3'
aP2	5'-TGATGCCTTTGTGGGAACCT-3'	5'-GCAAAGCCCCTCCACTT-3'
FAS	5'-GGACATGGTTCACAGACGATGAC-3'	5'-GTGCAACTTGGACAGATCCTTCA-3'

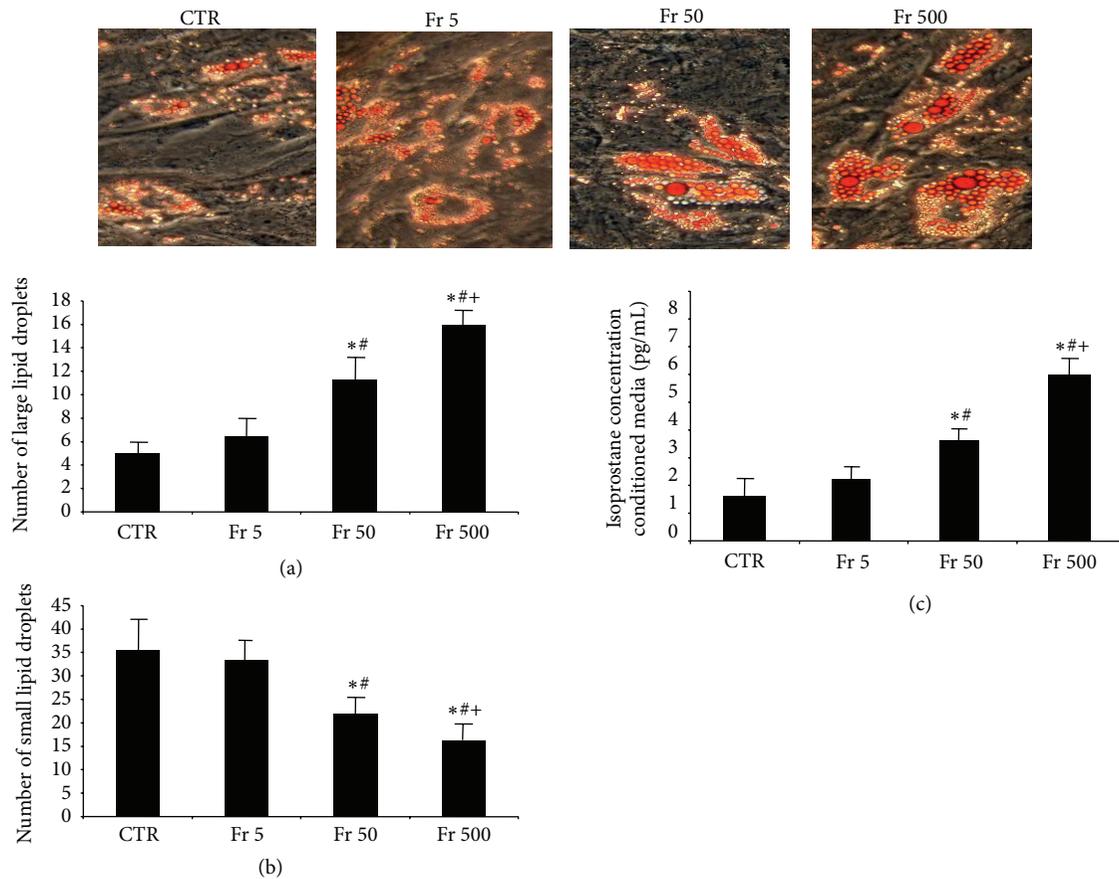


FIGURE 1: (a) Number of large lipid droplets from Oil Red O stained MSCs with 0, 5, 50, or 500 μ M of fructose, magnifications: 40x ($n = 3$), and (b) number of small lipid droplets from Oil Red O stained MSCs with 0, 5, 50, or 500 μ M of fructose, magnifications: 40x ($n = 3$). A representative section for each group is shown. (c) Measurement of isoprostane levels in conditioned media of differentiating MSCs. Values represent means \pm SEM of five independent treatments. * $P < 0.05$ versus control or # $P < 0.05$ versus Fr 5 μ M or + $P < 0.05$ versus Fr 50 μ M.

overexpression of HO-1, we measured the expression of HO-1 and adiponectin in pelleted cells, along with isoprostane and IL-6 levels in collected conditioned media after 14 days. Fructose treatment decreased the levels of both HO-1 and adiponectin compared to control (Figures 4(a) and 4(b), $P < 0.05$). Parallel to this finding, isoprostane and IL-6 levels in cells treated with fructose increased compared to control (Figures 4(c) and 4(d), $P < 0.05$). CoPP produced an increase in HO-1 and adiponectin expression compared to fructose alone ($P < 0.05$). There was a concomitant

decrease in isoprostane and IL-6 levels compared to the fructose group ($P < 0.05$). SnMP reversed the effect of CoPP, suggesting a regulatory role of HO activity in attenuating fructose-mediated adipocyte damage. We then measured mRNA levels of the canonical Wnt signaling molecules β -catenin and Wnt10b, along with aP2 and Peg-1/Mest. Fructose decreased the expression of β -catenin and Wnt10b in pelleted cells compared to control (Figures 5(a) and 5(b), resp., $P < 0.05$) and increased the expression of aP2 and Peg-1/Mest compared to control (Figures 5(c) and 5(d), $P < 0.05$). CoPP

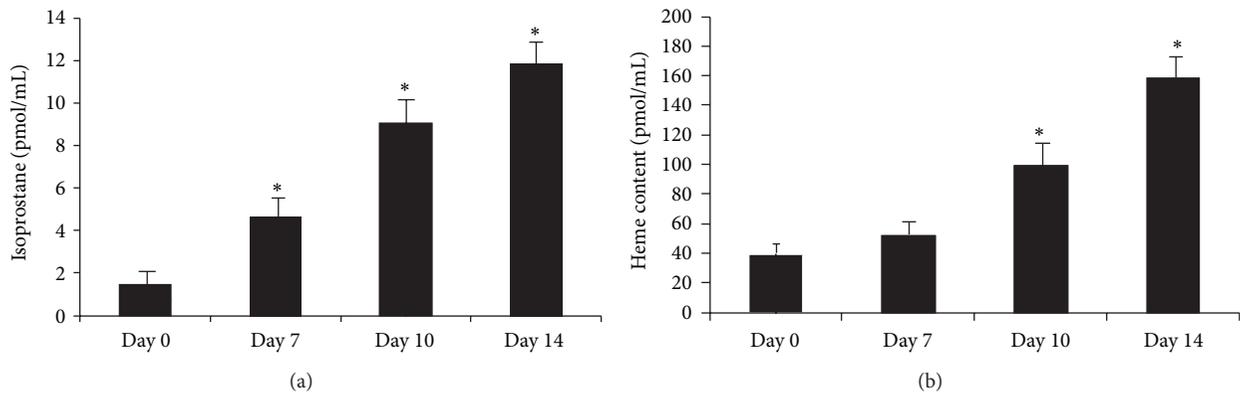


FIGURE 2: Measurement of 8-isoprostane in conditioned media and heme content in pelleted cells at days 0, 7, 10, and 14 of adipogenesis in human MSCs treated with 500 μM fructose (Figures 2(a) and 2(b), resp.). Values represent means ± SEM of five independent treatments. * $P < 0.05$ versus control.

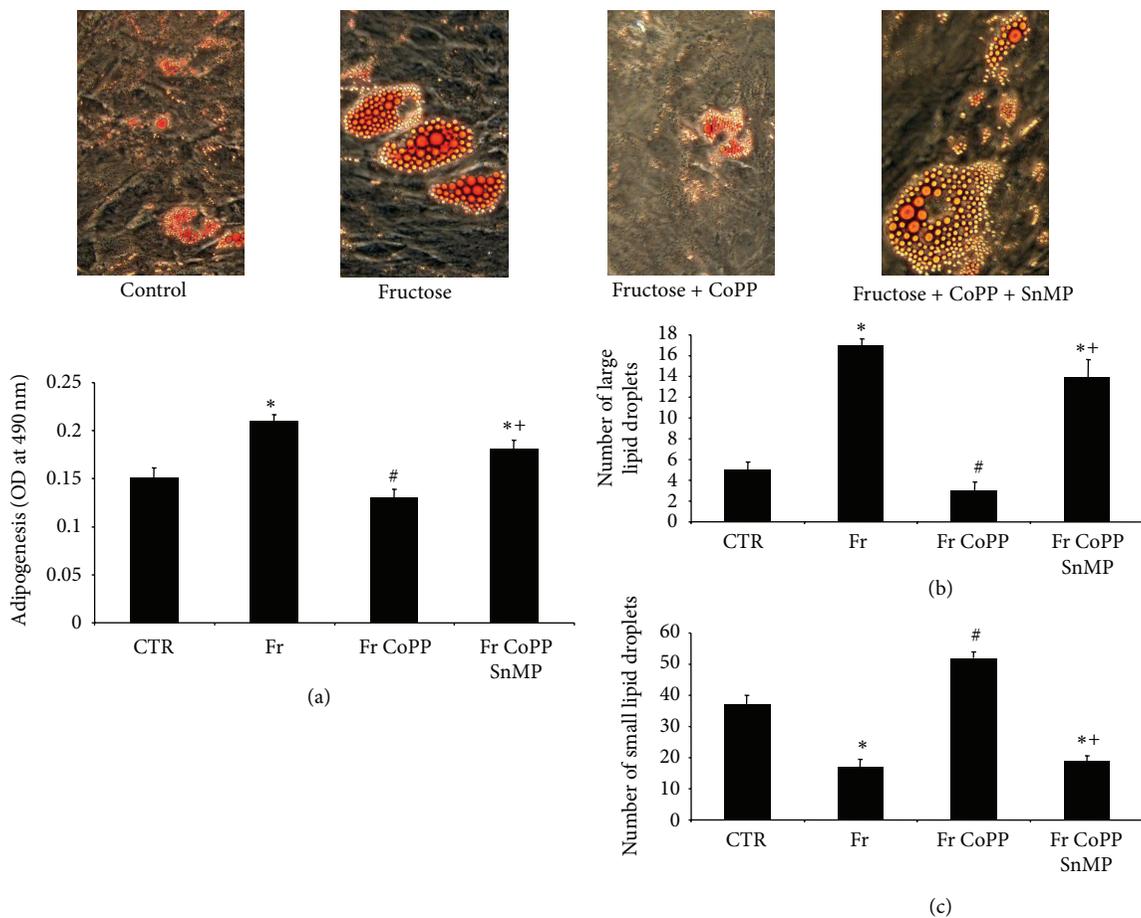


FIGURE 3: (a) Adipogenesis as shown by Oil Red O staining in MSCs treated with 500 μM fructose with or without CoPP, in the presence or absence of SnMP, magnifications and lipid droplet size from Oil Red O stained MSCs: 40x ($n = 3$). A representative section for each group is shown. (b) Number of large lipid droplets from Oil Red O stained MSCs treated with 500 μM fructose with or without CoPP, in the presence or absence of SnMP. (c) Number of small lipid droplets from Oil Red O treated with 500 μM fructose with or without CoPP, in the presence or absence of SnMP. Values represent means ± SEM of five independent treatments. * $P < 0.05$ versus control or # $P < 0.05$ versus fructose or ++ $P < 0.05$ versus fructose + CoPP.

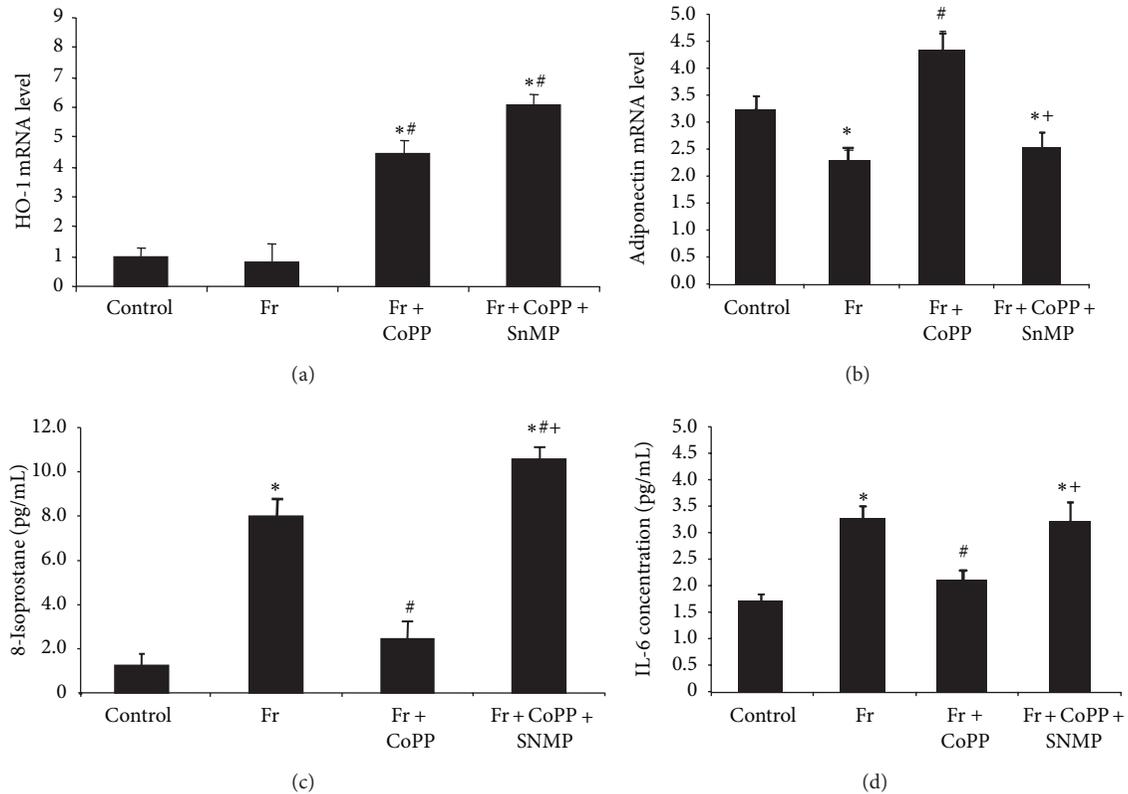


FIGURE 4: (a) Real-time PCR of HO-1 expression; (b) adiponectin expression; (c) concentration of 8-isoprostane in MSCs separated into control, fructose, fructose treated with CoPP, and fructose treated with CoPP and SnMP; and (d) concentration of IL-6 in conditioned media. Values represent means \pm SEM of five independent treatments. * $P < 0.05$ versus control or # $P < 0.05$ versus fructose or + $P < 0.05$ versus fructose with CoPP.

increased expression of β -catenin and Wnt10b compared to fructose treatment ($P < 0.05$) while simultaneously decreasing the expression of aP2 and Peg-1/Mest compared to the fructose group ($P < 0.05$). The beneficial effect of CoPP was reversed by concurrent treatment with SnMP ($P < 0.05$).

3.5. High Fructose Diet Increases Adipose Heme Content and Serum Isoprostane Levels In Vivo. Adipose tissue heme content increased compared to control in animals fed a high fructose diet (Figure 6, $P < 0.05$). CoPP ameliorated this increase ($P < 0.05$) and the concurrent administration of SnMP reversed the effects of CoPP ($P < 0.05$). Isoprostane levels in plasma were increased in animals fed a high fructose diet (Figure 7(a), $P < 0.05$ versus control); however, CoPP decreased isoprostane levels ($P < 0.05$); SnMP attenuated the beneficial effect of CoPP ($P < 0.05$). TNF α , IL-1, and IL-6 serum levels increased compared to control in mice fed a high fructose diet (Figures 7(b)–7(d), $P < 0.05$). CoPP ameliorated this increase in inflammatory cytokines ($P < 0.05$) to levels below control and the concurrent administration of SnMP reversed the effects of CoPP ($P < 0.05$). These findings support HO-1 as a key regulatory factor in reducing fructose-mediated increases in heme and isoprostane production *in vivo*.

3.6. HO-1 Overexpression Ameliorates Fructose-Mediated Adiposity and Adipocyte Dysfunction In Vivo. HO-1, adiponectin, and the adipogenic markers fatty acid synthase (FAS) and aP2 were measured in adipose tissue. A fructose diet decreased the levels of HO-1 and adiponectin compared to animals fed a normal chow diet (Figures 8(a) and 8(b), $P < 0.05$). Mice fed a high fructose diet exhibited a concurrent increase in FAS and aP2 gene expression in adipose tissue compared to mice fed a normal chow diet (Figures 8(c) and 8(d), $P < 0.05$). CoPP increased HO-1 and adiponectin expression compared to both the control and fructose groups ($P < 0.05$) and decreased the levels of FAS and aP2 compared to the fructose group ($P < 0.05$). In contrast, SnMP reversed the beneficial effects of CoPP ($P < 0.05$).

4. Discussion

This is the first study to show that upregulation of HO-1 gene expression decreased fructose-mediated isoprostane and heme levels in MSC-derived adipocytes and in mice fed a high fructose diet, thereby decreasing adipocyte dysfunction. These findings are associated with decreased inflammatory cytokine levels, increased adiponectin, and the elevated expression of genes of the canonical Wnt signaling cascade. Thus, the upregulation of HO-1 may provide a potential target

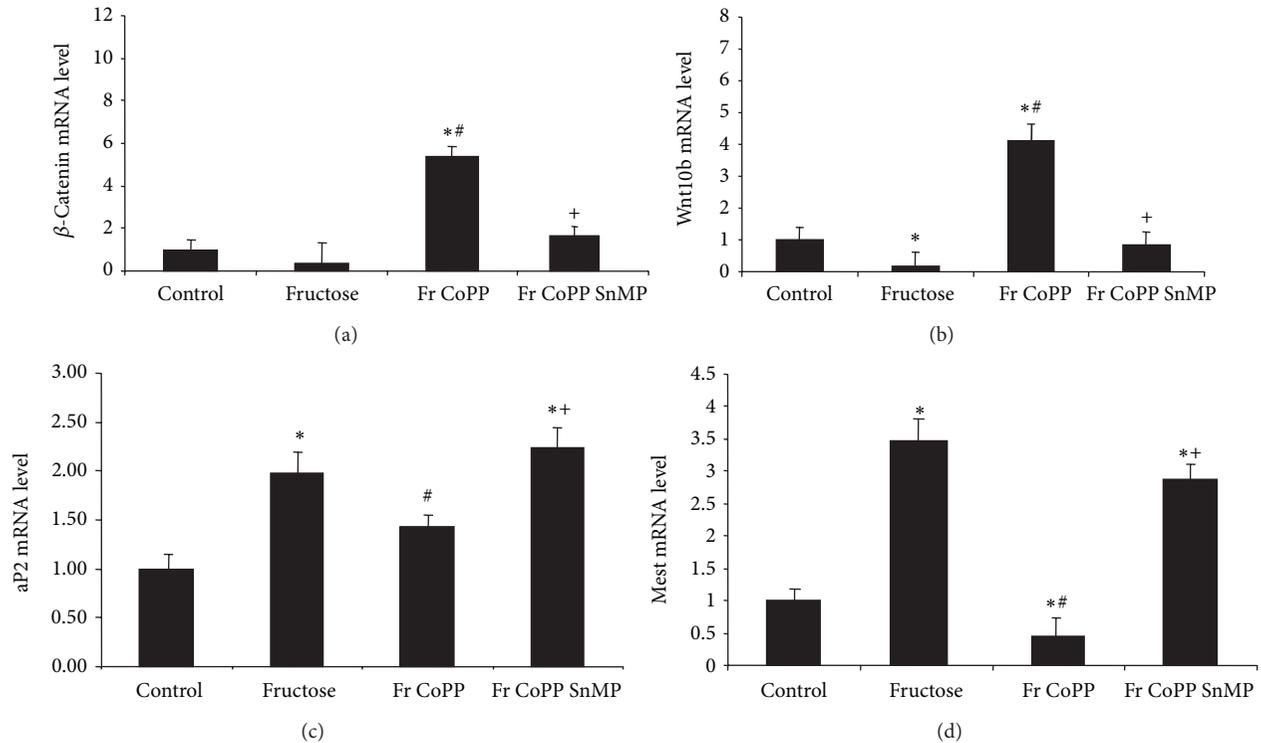


FIGURE 5: (a) Real-time PCR of β -catenin; (b) Wnt10b; (c) aP2; and (d) Peg-1/Mest expression in MSCs separated into control, fructose, fructose treated with CoPP, and fructose treated with CoPP and SnMP. Values represent means \pm SEM of five independent treatments. * P < 0.05 versus control or # P < 0.05 versus fructose or + P < 0.05 versus fructose with CoPP.

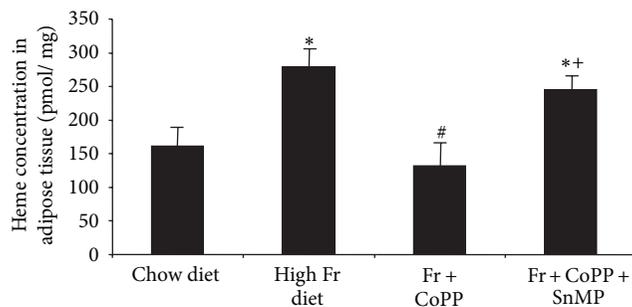


FIGURE 6: Cellular heme content from adipose tissue in mice fed chow diet, high fructose diet, and high fructose diet treated with CoPP, in the presence or absence of SnMP. Values represent means \pm SEM of four independent mice per group. * P < 0.05 versus chow diet or # P < 0.05 versus high fructose diet or + P < 0.05 versus high fructose with CoPP.

for pharmacological intervention in the treatment of patients with diet-induced obesity and metabolic syndrome without side effect. Mechanisms that govern metabolic adaptation to food intake during CoPP-mediated increase in HO-1 induced weight loss are not well understood. The loss of fat suggests that CoPP treated animals preferably used fatty acid as energy sources during weight loss. Weight loss with a normal appetite is a common specific response that is well documented [35–38].

The increased expression of HO-1 resulted in decreased isoprostane levels in fructose-mediated adiposity both *in vitro* and *in vivo*, whereas inhibition of HO activity, by SnMP, increased adipocyte differentiation and isoprostane production. The perturbations in isoprostane levels and adipogenesis as a result of modulation of HO-1 levels are in part related to adipose heme content. Heme, a known prooxidant, contributes to the generation of ROS [9] and increased amounts of heme cause stem cell differentiation and the release of inflammatory cytokines [26, 39–41]. In concordance with previous reports, our results show that the increase in cellular heme appears to constitute a major factor in the fructose-mediated isoprostane release, the increase in oxidative stress, and adipocyte dysfunction. Our results suggest that one of the mechanisms by which HO-1 modulation affects adiposity and decreases isoprostane levels is via the control of the cellular heme pool. We have previously shown that CoPP increased HO-1 protein expression in the adipocyte, in adipose tissue of human MSC-derived adipocytes, and in mice fed high fat diets [25, 42]. CoPP, through the induction of HO-1, attenuated the “molecular switch” of isoprostane and heme to genetically reprogram the adipocyte phenotype and prevent fructose-mediated adiposity and adipocyte dysfunction.

Fructose bypasses the two highly regulated steps of glycolysis, catalyzed by glucokinase/hexokinase and phosphofruktokinase. Instead, fructose enters the pathway at a level that is not regulated and excessive fructose metabolism leads to the direct activation of ROS through NADPH oxidase

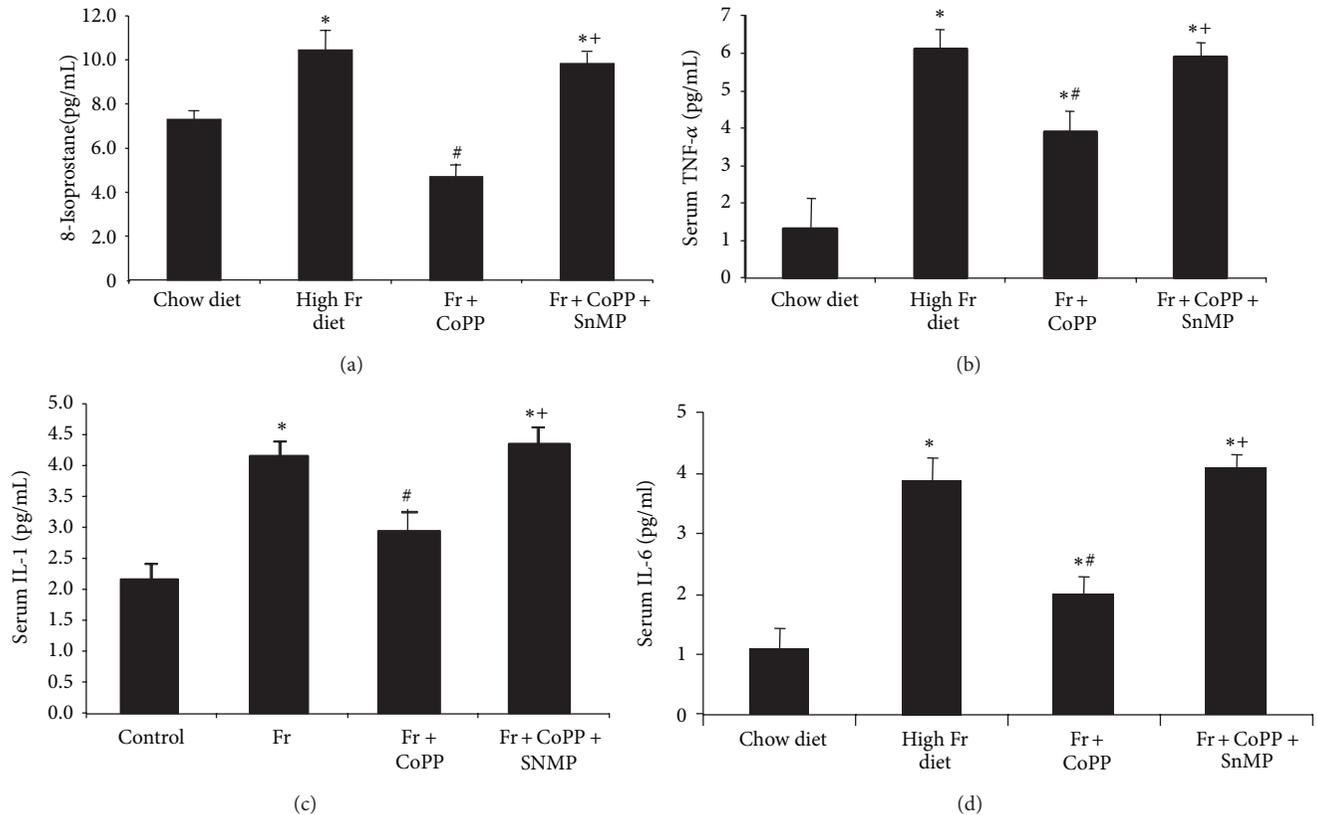


FIGURE 7: Assay of plasma concentrations of (a) isoprostane, (b) TNF α , (c) IL-1, and (d) IL-6 in mice fed chow diet or high fructose diet and high fructose diet treated with CoPP, in the presence or absence of SnMP. Values represent means \pm SEM of four independent mice per group. * $P < 0.05$ versus chow diet or # $P < 0.05$ versus high fructose diet or + $P < 0.05$ versus high fructose with CoPP.

[43]. Uncontrolled fructose metabolism leads to postprandial hypertriglyceridemia, which increases ROS. Previous findings have shown that obesity and ROS decreased HO-1 expression [24, 25]. Elevated glucose levels decrease HO-1 expression [44, 45] analogous to what is observed in humans and rodents with obesity. In concordance with these findings, our results show that MSC-derived adipocytes treated with fructose and mice fed a high fructose diet resulted in diminished HO-1 expression.

Further, our results show that isoprostane levels increased in a dose-dependent manner with fructose treatment in MSCs-derived adipocytes and in mice fed a high fructose diet. The increase in isoprostanes was associated with a decrease in HO-1 expression and an increase in heme levels. Isoprostanes and heme are regarded as valid markers of oxidative stress [8, 10, 46, 47]. Isoprostane production increases in experimental models of diabetes and metabolic syndrome and is ameliorated by the induction of HO-1 [8, 46, 48]. Polyphenol derived diets prevent vascular dysfunction and increase nitric oxide associated with arterial induction of HO-1 [32]. Here, we show, for the first time, that isoprostane production is increased in MSCs treated with fructose in a dose-dependent manner and in mice fed a high fructose diet and that CoPP reverses this effect thus suggesting a potential mechanism by which fructose may lead to adipocyte dysfunction in obesity.

Several key findings in this report offer potential insights into how increased HO-1 expression resulted in decreased levels of fructose-mediated adipogenesis and obesity. Firstly, our *in vitro* and *in vivo* studies show that an increase in HO-1 gene expression resulted in an increase in adiponectin levels in the presence of excessive fructose intake. In addition, upregulation of HO-1 decreased serum isoprostane and heme content, while simultaneously diminishing the levels of inflammatory cytokines. It is noteworthy that heme levels are increased during adipogenesis [26, 49] while HO-1 protein levels and HO activity are decreased in models of obesity and metabolic syndrome [24, 27, 28]. Another mechanism by which upregulation of HO-1 may decrease adiposity is related to an increase in the level of CO and bilirubin, the latter being a potent antioxidant. Bilirubin has been shown to scavenge ROS [50, 51] and play a role in the inhibition of NADPH oxidase [52] and protein kinase C (PKC) [53], which are key modulators of oxidant-induced adipocyte dysfunction. Inhibition of biliverdin reductase, the enzyme that converts biliverdin to bilirubin, and the consequent decrease in bilirubin production increase ROS 3-fold [54]. Therefore, increased levels of bilirubin may play a significant role in decreasing oxidants in adiposity.

Another potential mechanism via which upregulation of HO-1 decreases adipocyte hypertrophy may be the activation of the canonical Wnt pathway. HO-1 acts upstream of

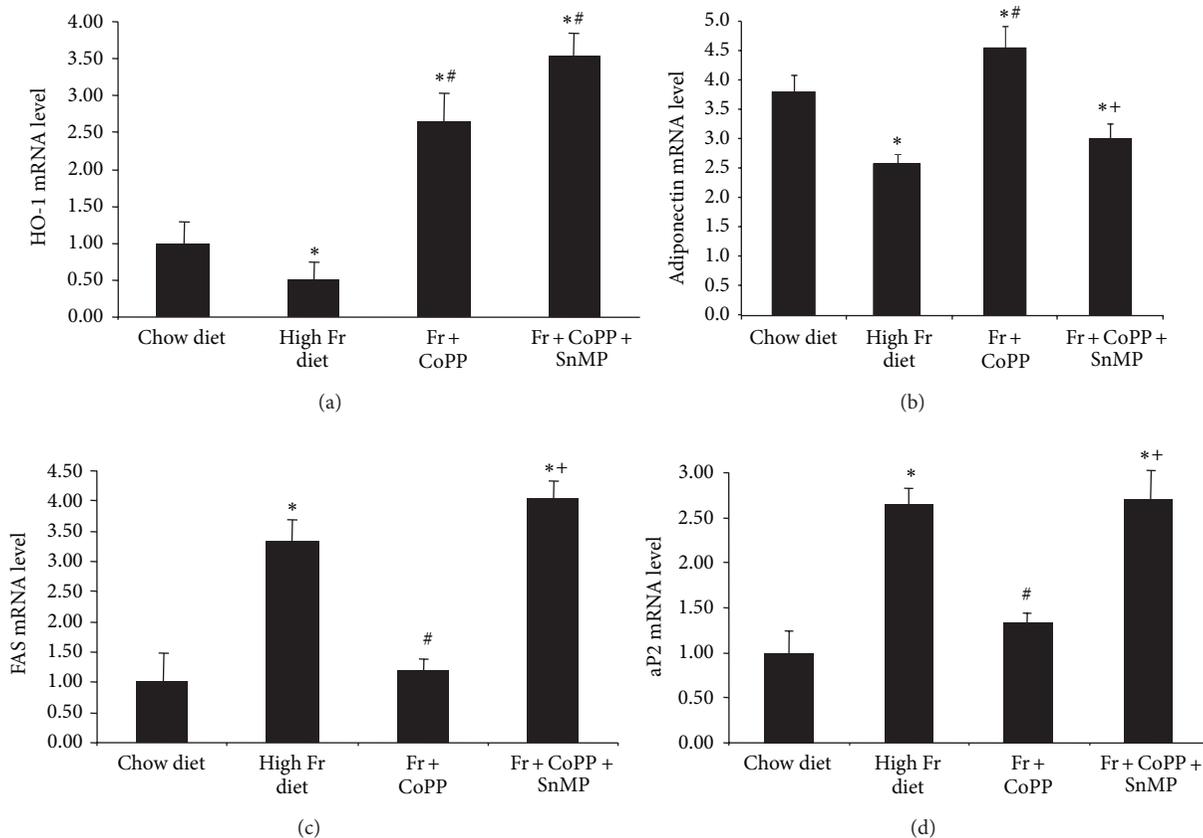


FIGURE 8: (a) Real-time PCR of HO-1, (b) adiponectin, (c) FAS, and (d) aP2 expression in chow diet, high fructose diet, and fructose diet treated with CoPP, in the presence or absence of SnMP. Values represent means \pm SEM of five independent treatments. * P < 0.05 versus chow diet or # P < 0.05 versus high fructose diet or + P < 0.05 versus high fructose with CoPP.

the canonical Wnt cascade and decreases lipogenesis and adipocyte differentiation in MSCs [55]. Wnts regulate gene expression through either the canonical (β -catenin dependent) or the noncanonical pathway (β -catenin independent) [22, 23]. An increase in Wnt/ β -catenin signaling inhibits the adipogenic transcription factors *C/EBP α* and *PPAR γ* [56, 57]. *C/EBP α* and *PPAR α* are central to growth arrest, that is, a prerequisite for adipocyte differentiation. Additionally, *Peg-1/Mest* upregulation results in the enlargement of adipocytes during adipose tissue expansion. On accumulation of triglycerides, *Peg-1/Mest* levels are increased with a parallel signal to preadipocytes to enlarge in order to accommodate more triglycerides [58]. Adipocyte enlargement is associated with an increase in the levels of inflammatory cytokines and oxidative stress [27, 30, 59].

Our results show that the increase in HO-1 gene expression resulted in an increase in the canonical Wnt signaling cascade and a decrease in *Peg-1/Mest* levels. A decrease in *Peg-1/Mest* is beneficial in the control of obesity, since upregulation of *Peg-1/Mest* occurs in obese adipose tissue in several models of obesity [60, 61]. Our data demonstrate that the overexpression of HO-1 was effective in suppressing adipocyte differentiation, as evidenced by an increase in the canonical Wnt cascade and a decrease in *Peg-1/Mest*. These effects were reversed by SnMP, further demonstrating

that the HO-1-mediated increase in negative regulators of adipogenesis and decrease in *Peg-1/Mest* resulted in the attenuation of adipogenesis and lipid accumulation. Taken together, these observations provide compelling evidence that the HO-1-mediated increase in Wnt signaling pathway modulates fructose-mediated adipogenesis.

A final mechanism by which CoPP-mediated upregulation of HO-1 and HO activity could decrease lipid accumulation is through elevated levels of adiponectin. A decrease in oxidative stress results in adipocytes secreting more adiponectin [27]. Thus, adiponectin may be considered a therapeutic target for HO-1 in reducing fructose-mediated adipocyte dysfunction and lipid accumulation, as previous findings have shown that induction of HO-1 reduces isoprostane and inflammatory cytokine levels through an increase in adiponectin [29]. In contrast, inhibition of HO activity decreased the expression of adiponectin confirming that adipocyte hypertrophy results in a diminished secretion of adiponectin [55]. Furthermore, increased heme levels, as a result of reduced HO-1 expression and HO activity, decrease adiponectin secretion from adipose tissue and increase adipocyte dysfunction and inflammation [26]. Similarly, our results show that upregulation of HO-1 in *in vitro* and *in vivo* models of high fructose consumption resulted in an increase in adiponectin levels along with a decrease in isoprostane and

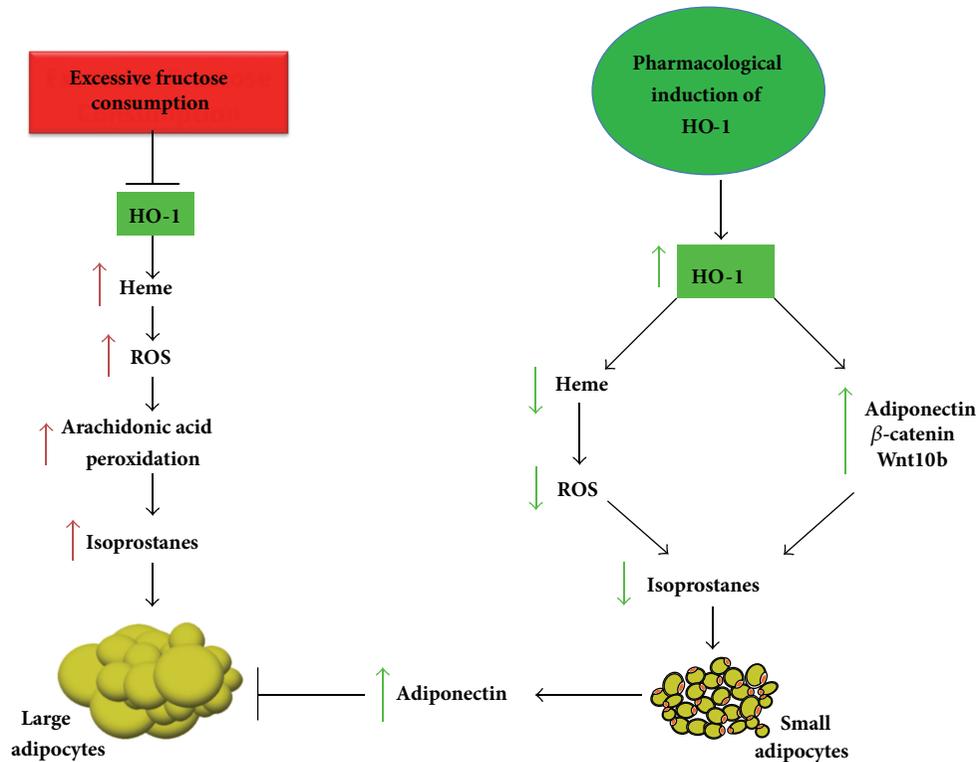


FIGURE 9: Schematic diagram of fructose-mediated adipocyte expansion and potential mechanisms underlying HO-1-mediated improvement of fructose-induced adipocyte hypertrophy. Excessive fructose consumption is accompanied by decreases in HO-1 gene expression and a concurrent increase in cellular heme levels. Increase in cellular heme results in lipid peroxidation of the essential fatty acid, arachidonic acid, via activation of ROS. The metabolite of this process is isoprostane and, therefore, heme catalyzes isoprostane synthesis. Isoprostane release induces adipocyte hypertrophy and dysfunction. In contrast, the increase in HO-1 expression and HO activity by pharmacological agents, for example, CoPP, leads to a decrease in the cellular heme pool and an increase in the canonical Wnt pathway and of adiponectin. Adiponectin enhances the antioxidant mechanisms, thereby decreasing lipid droplet size along with attenuation of isoprostane production. An increase in HO activity lowers lipid peroxidation, thereby decreasing isoprostane production and diminishing expression of the adipogenic markers Peg-1/Mest, FAS, and aP2, resulting in the lowering of adipocyte hypertrophy. Thus, activation of the HO-1-adiponectin axis along with an increase in the canonical Wnt cascade results in decreased heme (prooxidant) levels and isoprostane production in adipocytes and a decrease in fructose-mediated adipocyte dysfunction and lipid accumulation.

heme levels. Thus, activation of the HO-1-adiponectin axis may decrease fructose-mediated adipocyte dysfunction and metabolic imbalance.

In summary, this study demonstrates that the pharmacological induction of HO-1 results in a decrease in heme and isoprostane levels and a reduction in fructose-mediated adipocyte hypertrophy (as represented in Figure 9). We demonstrate that the stimulatory effect of exogenous fructose on isoprostane release exacerbates lipid accumulation and adipocyte dysfunction in MSC-derived adipocytes and mice fed a high fructose diet. Our results confirm the proadipogenic effects of fructose and account for the positive relationship between a high fructose diet and the presence of obesity. This may prove crucial in the control of obesity and the onset of metabolic syndrome as a result of a western high fructose diet.

What is already known about this subject is as follows.

- (i) Fructose consumption causes adiposity and increases reactive oxygen species (ROS) levels.

- (ii) Induction of heme oxygenase-1 (HO-1) decreases adiposity.

- (iii) Isoprostanes are a marker of oxidative stress.

What this study adds is as follows.

- (i) Fructose-mediated increase in adipogenesis is associated with an increase in isoprostane production in stem cell-derived adipocyte (MSCs).
- (ii) Fructose increases isoprostane production in a dose-dependent manner.
- (iii) Fructose increases the level of cellular heme, a prooxidant, in adipose tissue and the release of isoprostane into plasma.
- (iv) An increase in HO-1 expression reduces obesity and F2-isoprostane plasma levels in mice fed high fructose diets.

Abbreviations

aP2:	Adipocyte protein 2
β -Catenin:	Beta-cadherin associated protein
C/EBP α :	CCAAT/enhancer binding protein alpha
CoPP:	Cobalt protoporphyrin IX dichloride
FAS:	Fatty acid synthase
GLUT5:	Glucose transporter member 5
HO:	Heme oxygenase
HO-1:	Heme oxygenase isozyme 1, inducible form
HO-2:	Heme oxygenase isozyme 2, constitutive form
MSC:	Mesenchymal stem cell
NADPH:	Nicotinamide adenine dinucleotide phosphate (reduced form)
Peg-1/Mest:	Paternally expressed gene 1/mesoderm specific transcript
PPAR γ :	Peroxisome proliferator activated receptor gamma
ROS:	Reactive oxygen species
SnMP:	Tin mesoporphyrin IX dichloride
TNF α :	Tumor necrosis factor-alpha
Wnt10b:	Wingless-type MMTV integration site family, member 10b.

Disclosure

None of the authors have any financial, commercial, publication, or personal benefit.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

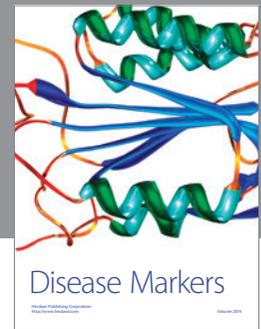
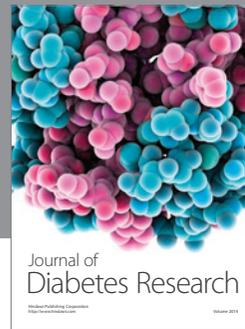
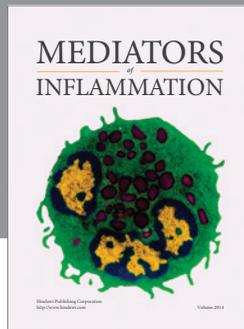
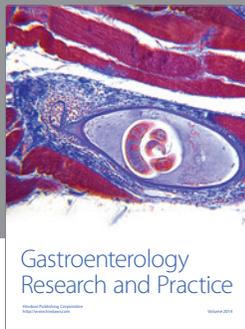
All authors had full access to the data and take responsibility for its integrity. All authors have read and agree with the paper as written. The authors thank Mrs. Jennifer Brown for secretarial support. This work was supported by National Institutes of Health Grants HL55601 and HL34300 (NGA) and HL109015 (JIS).

References

- [1] J. J. Rumessen and E. Gudmand-Hoyer, "Absorption capacity of fructose in healthy adults. Comparison with sucrose and its constituent monosaccharides," *Gut*, vol. 27, no. 10, pp. 1161–1168, 1986.
- [2] R. J. Johnson, S. E. Perez-Pozo, Y. Y. Sautin et al., "Hypothesis: could excessive fructose intake and uric acid cause type 2 diabetes?" *Endocrine Reviews*, vol. 30, no. 1, pp. 96–116, 2009.
- [3] R. J. Johnson, M. S. Segal, Y. Sautin et al., "Potential role of sugar (fructose) in the epidemic of hypertension, obesity and the metabolic syndrome, diabetes, kidney disease, and cardiovascular disease," *The American Journal of Clinical Nutrition*, vol. 86, no. 4, pp. 899–906, 2007.
- [4] T. Ishimoto, M. A. Lanaspas, M. T. Le et al., "Opposing effects of fructokinase C and A isoforms on fructose-induced metabolic syndrome in mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 11, pp. 4320–4325, 2012.
- [5] F. Q. Zhao and A. F. Keating, "Functional properties and genomics of glucose transporters," *Current Genomics*, vol. 8, no. 2, pp. 113–128, 2007.
- [6] K. L. Stanhope, J. M. Schwarz, N. L. Keim et al., "Consuming fructose-sweetened, not glucose-sweetened, beverages increases visceral adiposity and lipids and decreases insulin sensitivity in overweight/obese humans," *The Journal of Clinical Investigation*, vol. 119, no. 5, pp. 1322–1334, 2009.
- [7] N. G. Abraham and A. Kappas, "Pharmacological and clinical aspects of heme oxygenase," *Pharmacological Reviews*, vol. 60, no. 1, pp. 79–127, 2008.
- [8] D. Sacerdoti, R. Olszanecki, G. Li Volti, C. Colombrita, G. Scapagnini, and N. G. Abraham, "Heme oxygenase overexpression attenuates glucose-mediated oxidative stress in quiescent cell phase: linking heme to hyperglycemia complications," *Current Neurovascular Research*, vol. 2, no. 2, pp. 103–111, 2005.
- [9] J. Balla, H. S. Jacob, G. Balla, K. Nath, J. W. Eaton, and G. M. Vercellotti, "Endothelial-cell heme uptake from heme proteins: Induction of sensitization and desensitization to oxidant damage," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 20, pp. 9285–9289, 1993.
- [10] D. Sacerdoti, C. Colombrita, M. H. Ghattas et al., "Heme oxygenase-1 transduction in endothelial cells causes downregulation of monocyte chemoattractant protein-1 and of genes involved in inflammation and growth," *Cellular and Molecular Biology*, vol. 51, no. 4, pp. 363–370, 2005.
- [11] N. G. Abraham, T. Kushida, J. McClung et al., "Heme oxygenase-1 attenuates glucose-mediated cell growth arrest and apoptosis in human microvessel endothelial cells," *Circulation Research*, vol. 93, pp. 504–514, 2003.
- [12] T. Yura, M. Fukunaga, R. Khan, G. N. Nassar, K. F. Badr, and A. Montero, "Free-radical-generated F2-isoprostane stimulates cell proliferation and endothelin-1 expression on endothelial cells," *Kidney International*, vol. 56, no. 2, pp. 471–478, 1999.
- [13] J. D. Morrow and L. J. Roberts, "The isoprostanes: unique bioactive products of lipid peroxidation," *Progress in Lipid Research*, vol. 36, no. 1, pp. 1–21, 1997.
- [14] J. A. Awad, J.-L. Horn, L. J. Roberts II, and J. J. Franks, "Demonstration of halothane-induced hepatic lipid peroxidation in rats by quantification of F 2-isoprostanes," *Anesthesiology*, vol. 84, no. 4, pp. 910–916, 1996.
- [15] I. J. Tsai, K. D. Croft, T. A. Mori et al., "20-HETE and F2-isoprostanes in the metabolic syndrome: the effect of weight reduction," *Free Radical Biology and Medicine*, vol. 46, no. 2, pp. 263–270, 2009.
- [16] J. F. Keane Jr., M. G. Larson, R. S. Vasan et al., "Obesity and systemic oxidative stress: clinical correlates of oxidative stress in the Framingham study," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 23, no. 3, pp. 434–439, 2003.
- [17] P. Faure, C. Polge, D. Monneret, A. Favier, and S. Halimi, "Plasma 15-F2t isoprostane concentrations are increased during acute fructose loading in type 2 diabetes," *Diabetes & Metabolism*, vol. 34, no. 2, pp. 148–154, 2008.
- [18] M. Hokayem, E. Blond, H. Vidal et al., "Grape polyphenols prevent fructose-induced oxidative stress and insulin resistance in first-degree relatives of type 2 diabetic patients," *Diabetes Care*, vol. 36, no. 6, pp. 1454–1461, 2013.

- [19] Z. Khitan and D. H. Kim, "Fructose: a key factor in the development of metabolic syndrome and hypertension," *Journal of Nutrition and Metabolism*, vol. 2013, Article ID 682673, 12 pages, 2013.
- [20] P. Vestergaard, "Bone metabolism in type 2 diabetes and role of thiazolidinediones," *Current Opinion in Endocrinology, Diabetes and Obesity*, vol. 16, no. 2, pp. 125–131, 2009.
- [21] S. Gesta, Y.-H. Tseng, and C. R. Kahn, "Developmental origin of fat: tracking obesity to its source," *Cell*, vol. 131, no. 2, pp. 242–256, 2007.
- [22] H. Clevers, "Wnt/ β -catenin signaling in development and disease," *Cell*, vol. 127, no. 3, pp. 469–480, 2006.
- [23] K. M. Cadigan and Y. I. Liu, "Wnt signaling: complexity at the surface," *Journal of Cell Science*, vol. 119, no. 3, pp. 395–402, 2006.
- [24] J. Cao, S. J. Peterson, K. Sodhi et al., "Heme oxygenase gene targeting to adipocytes attenuates adiposity and vascular dysfunction in mice fed a high-fat diet," *Hypertension*, vol. 60, no. 2, pp. 467–475, 2012.
- [25] L. Vanella, D. H. Kim, K. Sodhi et al., "Crosstalk between EET and HO-1 downregulates Bach1 and adipogenic marker expression in mesenchymal stem cell derived adipocytes," *Prostaglandins and Other Lipid Mediators*, vol. 96, no. 1–4, pp. 54–62, 2011.
- [26] N. Puri, K. Sodhi, M. Haarstad et al., "Heme induced oxidative stress attenuates sirtuin1 and enhances adipogenesis in mesenchymal stem cells and mouse pre-adipocytes," *Journal of Cellular Biochemistry*, vol. 113, no. 6, pp. 1926–1935, 2012.
- [27] M. Li, D. H. Kim, P. L. Tsenovoy et al., "Treatment of obese diabetic mice with a heme oxygenase inducer reduces visceral and subcutaneous adiposity, increases adiponectin levels, and improves insulin sensitivity and glucose tolerance," *Diabetes*, vol. 57, no. 6, pp. 1526–1535, 2008.
- [28] S. J. Peterson, D. H. K. Kim, M. Li et al., "The L-4F mimetic peptide prevents insulin resistance through increased levels of HO-1, pAMPK, and pAKT in obese mice," *Journal of Lipid Research*, vol. 50, no. 7, pp. 1293–1304, 2009.
- [29] A. Nicolai, M. Li, D. H. Kim et al., "Heme oxygenase-1 induction remodels adipose tissue and improves insulin sensitivity in obesity-induced diabetic rats," *Hypertension*, vol. 53, no. 3, pp. 508–515, 2009.
- [30] S. J. Peterson, G. Drummond, D. H. Kim et al., "L-4F treatment reduces adiposity, increases adiponectin levels, and improves insulin sensitivity in obese mice," *Journal of Lipid Research*, vol. 49, no. 8, pp. 1658–1669, 2008.
- [31] A. L. Kruger, S. J. Peterson, M. L. Schwartzman et al., "Up-regulation of heme oxygenase provides vascular protection in an animal model of diabetes through its antioxidant and anti-apoptotic effects," *Journal of Pharmacology and Experimental Therapeutics*, vol. 319, no. 3, pp. 1144–1152, 2006.
- [32] Y. Shen, N. C. Ward, J. M. Hodgson et al., "Dietary quercetin attenuates oxidant-induced endothelial dysfunction and atherosclerosis in apolipoprotein E knockout mice fed a high-fat diet: a critical role for heme oxygenase-1," *Free Radical Biology & Medicine*, vol. 65, pp. 908–915, 2013.
- [33] J.-L. da Silva, M. Tiefenthaler, E. Park et al., "Tin-mediated heme oxygenase gene activation and cytochrome P450 arachidonate hydroxylase inhibition in spontaneously hypertensive rats," *American Journal of the Medical Sciences*, vol. 307, no. 3, pp. 173–181, 1994.
- [34] N. G. Abraham, J.-M. Camadro, S. T. Hoffstein, and R. D. Levere, "Effects of iron deficiency and chronic iron overloading on mitochondrial heme biosynthetic enzymes in rat liver," *Biochimica et Biophysica Acta—Protein Structure and Molecular Enzymology*, vol. 870, no. 2, pp. 339–349, 1986.
- [35] B. M. Grossman, M. L. Devore, E. W. Kelso, and R. J. Martin, "Effect of glucose and 2-deoxyglucose on hypothalamic GABA release in lactating rats," *Physiology & Behavior*, vol. 61, no. 2, pp. 169–173, 1997.
- [36] E. Scharrer, "Control of food intake by fatty acid oxidation and ketogenesis," *Nutrition*, vol. 15, no. 9, pp. 704–714, 1999.
- [37] M. A. Rupnick, D. Panigrahy, C.-Y. Zhang et al., "Adipose tissue mass can be regulated through the vasculature," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 16, pp. 10730–10735, 2002.
- [38] M. Morimura, O. Ishiko, T. Sumi, H. Yoshida, and S. Ogita, "Angiogenesis in adipose tissues and skeletal muscles with rebound weight-gain after diet-restriction in rabbits," *International Journal of Molecular Medicine*, vol. 8, no. 5, pp. 499–503, 2001.
- [39] S. R. Datla, G. J. Dusting, T. A. Mori, C. J. Taylor, K. D. Croft, and F. Jiang, "Induction of heme oxygenase-1 in vivo suppresses NADPH oxidase-derived oxidative stress," *Hypertension*, vol. 50, no. 4, pp. 636–642, 2007.
- [40] T. Chabrashvili, C. Kitiyakara, J. Blau et al., "Effects of ANG II type 1 and 2 receptors on oxidative stress, renal NADPH oxidase, and SOD expression," *The American Journal of Physiology—Regulatory Integrative and Comparative Physiology*, vol. 285, no. 1, pp. R117–R124, 2003.
- [41] K. Sodhi, N. Puri, D. H. Kim et al., "PPAR δ binding to heme oxygenase 1 promoter prevents angiotensin II-induced adipocyte dysfunction in Goldblatt hypertensive rats," *International Journal of Obesity*, vol. 38, no. 3, pp. 456–465, 2014.
- [42] A. Burgess, M. Li, L. Vanella et al., "Adipocyte heme oxygenase-1 induction attenuates metabolic syndrome in both male and female obese mice," *Hypertension*, vol. 56, no. 6, pp. 1124–1130, 2010.
- [43] K. L. Stanhope and P. J. Havel, "Fructose consumption: potential mechanisms for its effects to increase visceral adiposity and induce dyslipidemia and insulin resistance," *Current Opinion in Lipidology*, vol. 19, no. 1, pp. 16–24, 2008.
- [44] N. G. Abraham, T. Kushida, J. McClung et al., "Heme oxygenase-1 attenuates glucose-mediated cell growth arrest and apoptosis in human microvessel endothelial cells," *Circulation Research*, vol. 93, no. 6, pp. 507–514, 2003.
- [45] A. L. Kruger, S. Peterson, S. Turkseven et al., "D-4F induces heme oxygenase-1 and extracellular superoxide dismutase, decreases endothelial cell sloughing, and improves vascular reactivity in rat model of diabetes," *Circulation*, vol. 111, no. 23, pp. 3126–3134, 2005.
- [46] N. G. Abraham, "Heme oxygenase attenuated angiotensin II-mediated increase in cyclooxygenase activity and decreased isoprostane F 2α in endothelial cells," *Thrombosis Research*, vol. 110, no. 5–6, pp. 305–309, 2003.
- [47] C. Colombrita, G. Lombardo, G. Scapagnini, and N. G. Abraham, "Heme oxygenase-1 expression levels are cell cycle dependent," *Biochemical and Biophysical Research Communications*, vol. 308, no. 4, pp. 1001–1008, 2003.
- [48] N. G. Abraham, R. Rezzani, L. Rodella et al., "Overexpression of human heme oxygenase-1 attenuates endothelial cell sloughing in experimental diabetes," *The American Journal of Physiology—Heart and Circulatory Physiology*, vol. 287, no. 6, pp. H2468–H2477, 2004.

- [49] N. Kumar, L. A. Solt, Y. Wang et al., "Regulation of adipogenesis by natural and synthetic REV-ERB ligands," *Endocrinology*, vol. 151, no. 7, pp. 3015–3025, 2010.
- [50] J. E. Clark, R. Foresti, C. J. Green, and R. Motterlini, "Dynamics of haem oxygenase-1 expression and bilirubin production in cellular protection against oxidative stress," *Biochemical Journal*, vol. 348, no. 3, pp. 615–619, 2000.
- [51] J. Neuzil and R. Stocker, "Free and albumin-bound bilirubin are efficient co-antioxidants for α -tocopherol, inhibiting plasma and low density lipoprotein lipid peroxidation," *The Journal of Biological Chemistry*, vol. 269, no. 24, pp. 16712–16719, 1994.
- [52] J. Y. Kwak, K. Takeshige, B. S. Cheung, and S. Minakami, "Bilirubin inhibits the activation of superoxide-producing NADPH oxidase in a neutrophil cell-free system," *Biochimica et Biophysica Acta—Protein Structure and Molecular Enzymology*, vol. 1076, no. 3, pp. 369–373, 1991.
- [53] K. Sano, H. Nakamura, and T. Matsuo, "Mode of inhibitory action of bilirubin on protein kinase C," *Pediatric Research*, vol. 19, no. 6, pp. 587–590, 1985.
- [54] D. E. Barañano, M. Rao, C. D. Ferris, and S. H. Snyder, "Biliverdin reductase: a major physiologic cytoprotectant," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 25, pp. 16093–16098, 2002.
- [55] L. Vanella, K. Sodhi, D. H. Kim et al., "Increased heme-oxygenase 1 expression in mesenchymal stem cell-derived adipocytes decreases differentiation and lipid accumulation via upregulation of the canonical Wnt signaling cascade," *Stem Cell Research and Therapy*, vol. 4, no. 2, article 28, 2013.
- [56] M. Kawai, S. Mushiake, K. Bessho et al., "Wnt/Lrp/ β -catenin signaling suppresses adipogenesis by inhibiting mutual activation of PPAR γ and C/EBP α ," *Biochemical and Biophysical Research Communications*, vol. 363, no. 2, pp. 276–282, 2007.
- [57] M. Okamura, H. Kudo, K.-I. Wakabayashi et al., "COUP-TFII acts downstream of Wnt/ β -catenin signal to silence PPAR γ gene expression and repress adipogenesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 14, pp. 5819–5824, 2009.
- [58] M. Takahashi, Y. Kamei, and O. Ezaki, "Mest/Peg1 imprinted gene enlarges adipocytes and is a marker of adipocyte size," *The American Journal of Physiology—Endocrinology and Metabolism*, vol. 288, no. 1, pp. E117–E124, 2005.
- [59] L. Vanella, D. H. Kim, D. Asprinio et al., "HO-1 expression increases mesenchymal stem cell-derived osteoblasts but decreases adipocyte lineage," *Bone*, vol. 46, no. 1, pp. 236–243, 2010.
- [60] Y. Kamei, T. Suganami, T. Kohda et al., "Peg1/Mest in obese adipose tissue is expressed from the paternal allele in an isoform-specific manner," *FEBS Letters*, vol. 581, no. 1, pp. 91–96, 2007.
- [61] R. A. Koza, L. Nikonova, J. Hogan et al., "Changes in gene expression foreshadow diet-induced obesity in genetically identical mice," *PLoS Genetics*, vol. 2, no. 5, article e81, 2006.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

