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Role of Serum Biomarkers in Early Detection of Diabetic Cardiomyopathy in the West Virginian Population

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Abstract

Objectives: Diabetic cardiomyopathy (DCM) is an established complication of diabetes mellitus. In West Virginia, the especially high incidence of diabetes and heart failure validate the necessity of developing new strategies for earlier detection of DCM. Since most DCM patients remain asymptomatic until the later stages of the disease when the fibrotic complications become irreversible, we aimed to explore biomarkers that can identify early-stage DCM. Methods: The patients were grouped into 4 categories based on clinical diabetic and cardiac parameters: Control, Diabetes (DM), Diastolic dysfunction (DD), and Diabetes with diastolic dysfunction (DM+DD), the last group being the preclinical DCM group. Results: Echocardiography images indicated severe diastolic dysfunction in patients with DD+DM and DD compared to DM or control patients. In the DM and DM+DD groups, TNFα, isoprostane, and leptin were elevated compared to control (p<0.05), as were clinical markers HDL, glucose and hemoglobin A1C. Fibrotic markers IGFBP7 and TGF-β followed the same trend. The Control group showed higher beneficial levels of adiponectin and bilirubin, which were reduced in the DM and DM+DD groups (p<0.05). Conclusion: The results from our study support the clinical application of biomarkers in diagnosing early stage DCM, which will enable attenuation of disease progression prior to the onset of irreversible complications.

Key words: cardiomyopathy, diabetes, West Virginia, serum biomarkers.

Introduction

In recent years, diabetes mellitus (DM) has become a national health epidemic. The Centers for Disease Control and Prevention reports that West Virginia has one of the highest rates of diabetes in the United States with more than 11% of the population affected. The Framingham Heart Study revealed that the risk of heart failure is up to 5 times higher in diabetics than non-diabetics, when controlling for other risk factors. Diabetic Cardiomyopathy (DCM) is an established complication of diabetes [1-6] that involves abnormal relaxation of the ventricles, referred to as diastolic dysfunction, with concurrent hypertrophy of cardiomyocytes [6, 7]. Diastolic dysfunction is thought to be the first functional abnormality in DCM and can be seen in 40-60% of asymptomatic diabetic patients through echocardiographic imagining studies [2, 6]. Diabetic patients with subclinical diastolic dysfunction have a 5-year mortality rate of 30.8%
compared to 12.1% for diabetic patients with no diastolic dysfunction [4]. As DCM enters its later stage, it progresses from diastolic dysfunction to overt stage C heart failure with preserved ejection fraction, which has no proven effective treatment [7], thus validating the importance of identifying biomarkers that can improve detection of DCM prior to the onset of irreversible complications.

Diabetes impairs glucose uptake and results in an increase in fatty acid (FA) metabolism in cardiac tissue [3, 8, 9]. In diabetes, decreased insulin signaling activates transcriptional signaling pathways that induce the expression of genes involved in stimulating FA uptake; however the uptake of FAs exceeds metabolic demand and results in triglyceride and cholesterol accumulation in the myocardium which impairs diastolic function [8-11]. A study by McGavock et al compared normoglycemic individuals with diabetic patients and confirmed a positive correlation between impaired glucose tolerance and myocardial triglyceride content and found that triglyceride accumulation preceded the onset of ventricular dysfunction [11]. Abnormal FA metabolism also leads to depressed levels of high-density lipoprotein (HDL) [3].

Multiple studies have established a link between damage induced by oxidative stress and DCM [12, 13]. Damage from oxidative stress due to the chronic mitochondrial overproduction of reactive oxygen species (ROS) plays a crucial role in inflammation and results in irreversible fibrosis and cardiomyocyte death [2, 12, 14, 15]. Inflammation in the myocardium is mediated by pro-inflammatory cytokines including TNFα and interleukin-6 [16]. Isoprostanes are formed by the peroxidation of polyunsaturated FAs and are considered an accurate reflection of the extent of oxidative damage [17]. Amelioration of oxidative stress on a molecular level can be achieved through induction of antioxidant agents, and studies have shown that enhancing mitochondrial ROS scavenging systems mitigates diabetes-induced cardiac dysfunction [2, 12, 18-21]. Bilirubin, a product of heme catabolism, is a potent antioxidant and under normal physiological conditions may attenuate many ROS-derived complications of DCM [22, 23]. Adiponectin is a hormone secreted by adipose tissue that regulates metabolic processes and functions as an antioxidant; the low plasma levels of adiponectin seen in diabetes contribute to the oxidative damage seen in DCM [24, 25].

Structurally, the progression of DCM has been linked to cardiomyocyte hypertrophy and increased fibrosis [26-29]. The presence of cardiomyocyte hypertrophy was supported by data from The Framingham Heart Study, which revealed left ventricular mass was higher in diabetics compared to non-diabetics independent of covariates [30]. Hyperglycemia facilitates the reaction of glucose with collagen to form advanced glycation end-products (AGEs) that promote the crosslinking of collagen molecules to produce fibrosis [26]. Insulin-like growth factor binding protein 7 (IGFBP7) is a modulator of insulin-like growth factors, which actively regulate insulin consumption and receptor activity [27]. IGFBP7 has been identified as a biomarker for diabetic dysfunction associated with cardiac hypertrophy, myocyte fibrosis, and vascular remodeling, and an association exists between serum IGFBP7 levels and echocardiographic abnormalities describing diastolic heart dysfunction independent of systolic function [28]. In diabetics, increased ROS production causes the subsequent release of the profibrotic marker transforming growth factor beta (TGF-β) [29]. TGF-β levels correlate with the degree of cardiac fibrosis [26]. The reliable early detection of fibrotic changes in DCM patients by IGFBP7 and TGF-β is clinically pertinent since current experimental therapies designed to prevent progression of DCM include targeting collagen deposition and fibrosis through AGE inhibitors and AGE cross-link breakers [1, 6].

The aim of this study was to investigate a panel of biomarkers, including fibrotic markers, antioxidants, markers of oxidative stress and a metabolic profile that can be used to detect DCM prior to the presentation of clinical signs. Early stage DCM is initially asymptomatic and reversible; however, DCM is a progressive disease and at the later stage prognosis becomes poor. Analysis of biomarkers would be a simple, noninvasive diagnostic tool with a high predictive value for clinical use in patients with the potential to develop DCM, which would allow for the implementation of therapeutic strategies to be started in time to delay and mitigate the disease progression.

Materials and Methods

Patients

This study comprised of 100 patients, divided into 4 categories based on predetermined clinical diabetic and cardiac parameters: 1. Control; 2. Diabetes with normal diastolic function; 3. Diastolic dysfunction without diabetes; 4. Diabetes and diastolic dysfunction. Contraindications to inclusion included any systolic dysfunction observed through echocardiography, evidence of prior myocardial infarction, known valvular or congenital heart disease or other significant comorbidities, including malignancy, renal failure, or significant psychiatric illness. The study was approved by the Ethics Committee of Cabell Huntington Hospital, West Virginia. All of the subjects signed a written informed consent form.
Table 1: Demographic Profile

<table>
<thead>
<tr>
<th>Groups</th>
<th>Control</th>
<th>Diastolic dysfunction (DD)</th>
<th>Diabetes mellitus (DM)</th>
<th>Diabetes mellitus with diastolic dysfunction (DM+DD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yrs.)</td>
<td>41.67±11.79</td>
<td>44.27±8.31</td>
<td>48±11.78</td>
<td>45.69±8.42</td>
</tr>
<tr>
<td>Sex (M/F)</td>
<td>43%/57%</td>
<td>50%/50%</td>
<td>52%/48%</td>
<td>57%/43%</td>
</tr>
</tbody>
</table>

Blood Samples

Venous blood was drawn from an antecubital vein in heparinized and Vacutainer tubes for serum levels of TNFα, IL-6, Isoprostane, leptin, adiponectin, bilirubin, IGFBP7, TGF-β and lipid and glucose profiles. Blood was processed after collection and serum collected for analysis. Serum was collected by spinning down the vacutainer tube at 2000 rpm for 5 minutes at 4°C and stored at -80°C until analysis.

Analysis of Biomarkers

Serum samples frozen at -80°C were analyzed for levels of the following biomarkers through enzyme-linked immunosorbent-assay (ELISA): TNFα, IL-6, Isoprostane, leptin, IGFBP7, TGF-β and lipid and glucose profiles. Blood was processed after collection and serum collected for analysis. Serum was collected by spinning down the vacutainer tube at 2000 rpm for 5 minutes at 4°C and stored at -80°C until analysis.

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Echocardiography

Patients underwent echocardiography with 2D, Doppler, and color flow imaging within the year prior to enrollment. Studies were performed on a Philips IE 33 with an S5 transducer by one of 2 certified echo technicians in an ICAEL accredited laboratory. Measurements were obtained in accordance with guidelines set forth by the American Society of Echocardiography [31]. 2D imaging was used to assess ejection fraction (EF), septal wall thickness, posterior wall thickness and left atrial size. Relative wall thickness was calculated with posterior and septal wall thickness by (2×PWTd)/LVIDd [32]. Mitral valve inflow Doppler was recorded, including E and A waves. Tissue Doppler was used for E’ and A’. Echocardiographic images were read by physicians who were blinded to the rest of the study.

Statistical Analysis

Data were analyzed using Graphpad Prism 4.0. Equal variance was assured by Bartlett’s test for each biomarker within each of the four patient categories. ANOVA was conducted to identify statistically significant differences in the mean serum levels for the different biomarkers. The Tukey post-hoc test was used to indicate which patient groups showed statistically significant differences for the biomarker level measured.

Results

Echocardiography

All patients underwent transthoracic echocardiography to evaluate left ventricular diastolic filling pressure. Patients were identified as normal or as having diastolic dysfunction (DD), with DD encompassing a spectrum of severities including impaired relaxation, pseudonormal and restrictive diastolic dysfunction (Fig. 1). Results from echocardiographic imaging studies were used in conjunction with the diabetic status of the patients to establish the 4 patient categories.

TNFα, isoprostane and IL-6 levels

TNFα, isoprostanes, and IL-6 are inflammatory mediators that correlate with the extent of oxidative damage in tissues. TNFα and IL-6 were found to be elevated in DM, DD and DM+DD patients when compared to the control group (Fig. 2A and B respectively; p < 0.05). Levels of TNFα and IL-6 were significantly higher in patients with DM+DD than the levels found in either the patients with DM or the patients with DD alone (p < 0.05). Patients with DM and patients with DM+DD had increased levels of isoprostane compared to the control (Fig. 2C). Additionally, isoprostane levels in patients with DM+DD were higher than levels observed in patients with DD alone (p < 0.05).

Table 2: Patient Clinical Profile.

<table>
<thead>
<tr>
<th></th>
<th>BMI (±1.01)</th>
<th>Triglycerides (mg/dL) (±11.02)</th>
<th>HDL (mg/dL) (±3.96)</th>
<th>Glucose (mg/dL) (±4.50)</th>
<th>Hba1C (mg/dL) (±0.11)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>23.89</td>
<td>61.12</td>
<td>67.60</td>
<td>91.50</td>
<td>5.20</td>
</tr>
<tr>
<td>DM</td>
<td>39.79*</td>
<td>85.72</td>
<td>43.60*</td>
<td>186.28*</td>
<td>8.04*</td>
</tr>
<tr>
<td>DD</td>
<td>31.42*</td>
<td>92.16</td>
<td>44.42*</td>
<td>107.23*</td>
<td>5.72*</td>
</tr>
<tr>
<td>DM+DD</td>
<td>39.44*</td>
<td>113.05*</td>
<td>39.50*</td>
<td>172.9*</td>
<td>7.36*</td>
</tr>
</tbody>
</table>

Values represent means ± SEM. * p<0.05 vs Control; # p<0.05 vs DM; + p<0.05 vs DD.
Figure 1: Mitral inflow velocities representing normal physiology as well as the spectrum of diastolic dysfunction. A: Normal. B: Impaired Relaxation. C: Grade II diastolic dysfunction (pseudonormalization). D: Restrictive

Figure 2: Assay of serum concentrations of (A) TNFα, (B) IL-6 and (C) Isoprostane in Control, DM, DD and DM+DD patient groups. Values represent means ± SEM. * p<0.05 vs CTR; # p<0.05 vs DM, + p<0.05 vs DD.
Leptin, adiponectin and bilirubin levels

Leptin is an adipose-derived hormone that stimulates energy expenditure and is proportional to the amount of body fat. Leptin was significantly higher in the DM, DD and DM+DD patients when compared to the control group and was also significantly increased in the DM+DD patients when compared to the patients with DD alone (Fig. 3A; p < 0.05). Adiponectin was significantly lower in DM, DD, and DM+DD patients when compared to control groups (Fig. 3B). Adiponectin was also significantly decreased in DM+DD patients when compared to patients with DM and DD alone (p < 0.05). Bilirubin, a potent anti-oxidant, was found to be lowered in the DM+DD group compared to the control (Fig. 3C; p < 0.05).

TGF-β and IGFBP7 levels

TGF-β correlates to the extent of fibrosis and IGFBP7 has been linked to cardiomyocyte hypertrophy and fibrosis. In concordance with our hypothesis, TGF-β levels were elevated in DM, DD and DM+DD patients when compared to the control group (Fig. 4A; p < 0.05). DM+DD patients did have significantly higher TGF-β levels than the DM group alone. IGFBP7 was found to be increased in the DM, DD and DM+DD groups compared to the control. IGFBP7 was also higher in patients with DM+DD compared to patients with DM or DD alone (Fig. 4B; p < 0.05).

Figure 3: Serum levels of (A) Leptin, (B) Adiponectin and (C) Bilirubin in Control, DM, DD and DM+DD patient groups determined by ELISA. Values represent means ± SEM. * p<0.05 vs CTR; # p<0.05 vs DM, + p<0.05 vs DD.

Figure 4: Assay of serum concentrations of (A) TGF-β and (B) IGFBP7 in Control, DM, DD and DM+DD patient groups. Values represent means ± SEM. * p<0.05 vs CTR; # p<0.05 vs DM, + p<0.05 vs DD.
Discussion

This study demonstrated that a panel of biomarkers in conjunction with echocardiography plays an important role in the detection of DCM prior to the manifestation of symptoms and irreversible complications. Our data indicated that early stage DCM can be characterized by elevated levels of inflammatory markers, decreased antioxidant markers and, most importantly, increased levels of the fibrotic markers IGFBP7 and TGF-β. These results, which are summarized in the schematic diagram (Fig. 5), identified potential biomarkers that can be used clinically to improve early diagnosis of DCM.

The clinical parameters studied include BMI and levels of triglycerides, HDL, glucose and HbA1C. A comprehensive national study, which substantiates the correlation between BMI and diabetes, found that individuals with diabetes were more likely to have a high BMI [33]. In concordance with established studies, our results demonstrated elevated blood glucose levels and HbA1C levels in both DM patients and patients with DM+DD. As expected, neither glucose nor HbA1C was elevated in DD patients, because DD is not specific to DCM but can result from hypertension, aortic stenosis, myocardial infarction, or systolic heart failure [31]. Triglycerides are elevated in diabetic patients due to an increased reliance on fatty acid metabolism, and our data demonstrated DM+DD patients had significantly elevated triglyceride levels. Additionally, our data supported the accepted trend that diabetes is closely associated with reduced levels of HDL.

Elevated levels of the inflammatory cytokines TNFα and IL-6 are associated with diabetes, and previous studies have indicated their role in cardiac remodeling, fibrosis and cardiomyocyte apoptosis [34]. In corroborations with these findings, our study demonstrated elevated levels of TNFα and IL-6 relative to the control in DM, DD, and DM+DD patients, with the highest levels in DM+DD patients, supporting their utilization in the early detection of DCM and amelioration of disease progression. Isoprostane, an established marker of oxidative stress [11], was increased in patients with DM+DD. Our results also demonstrate a decreased level of the antioxidant bilirubin in patients with DM+DD, which supports previous findings that bilirubin is negatively associated with diabetes and its complications.

Leptin is closely associated with obesity and its comorbidities [35], and was highest in patients with DM and patients with DM+DD, which were the groups with the highest BMIs. There is an inverse relationship between adiponectin and leptin levels, with low adiponectin levels indicating metabolic and cardiovascular disease [36]. Adiponectin has antidiabetic effects and antagonizes the progression of DCM by exhibiting potent anti-hypertrophic activity in cardiomyocytes and diminishing cardiac cell death by inhibiting inflammation [36-38]. Consistent with previous studies, our data identified decreased levels of adiponectin in DM patients and DD patients and significantly lower adiponectin levels in DM+DD patients, which indicated a diminished ability to offset the DCM complications and progression.

IGFBP7 is an established marker for diabetes, and is associated with the severity of diastolic dysfunction [28]. IGFBP7 also has been implicated in fibrogenesis in diabetic patients, and increasing concentrations correlate with increased collagen deposition, which, in cardiac tissue, may contribute to diastolic stiffness [39]. Diabetes-induced ROS generation up-regulates the expression of the profibrotic marker TGF-β, leading to increased extracellular matrix synthesis and fibrosis [26]. Increased collagen deposition and interstitial fibrosis are key pathogenic components of DCM that result in myocardial stiffness, which manifests as diastolic dysfunction [2]. In concordance with previous studies, our data indicated elevated IGFBP7 and TGF-β levels in DM, DD and DM+DD patients, with DM+DD patients displaying the highest values. Therefore, these markers play an important role in the early detection of DCM and cardiac fibrosis, enabling early intervention to attenuate disease progression.

Figure 5: Schematic depicting the progression from diabetes to DCM and the role of biomarkers to detect early stage DCM and attenuate disease progression.
Conclusion

In conclusion, our study identified a panel of biomarkers that can complement echocardiographic imaging studies to detect the diabetes-induced changes in cardiac structure and function present at the early stage of DCM. The progression of DCM from subclinical diastolic dysfunction to overt heart failure, and the associated high morbidity and mortality in the later stage of the disease, validates the need for reliable early diagnostic tools. The results of our clinical study can improve early diagnosis of DCM and help successfully implement preventative strategies that may slow or prevent development of the disease.

Our study advances the understanding of the clinical use of biomarkers in the diagnosis of DCM in a rural Appalachian population. Diabetes is a chronic disease with high prevalence in West Virginia, and its comorbidities present a significant public health concern. Therefore, West Virginia patients present the ideal population to study strategies for earlier detection of DCM. Positive results from this study will benefit this underserved population by enabling an earlier and more reliable diagnosis of DCM so that therapy can be administered in time to attenuate disease progression.

Acknowledgement

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Competing Interests

The authors have declared that no competing interest exists.

References


